**Abstract**

Viral insect-borne plant pathogens have devastating impacts in agroecosystems. Vector-borne pathogens are often transmitted by generalist insects that move between non-crop and crop hosts. Insect vectors can have wide diet breadths, but it is often unknown which hosts serve as pathogen reservoirs and which non-crop host harbors the highest density of vectors. We addressed these issues by linking field surveys of an aphid vector and plant virus with statistical models to develop risk assessments for common non-crop legumes. In the Pacific Northwest USA, the pea aphid (*Acyrthosiphon pisum*) is a key virus vector with a wide host range; in 2018 we completed a 65-site survey where aphids were surveyed in weedy legumes within and outside dry pea fields. We quantified the abundance of pea aphids on seventeen hosts, and plant tissue was tested for pea enation mosaic virus (PEMV), a pathogen that causes considerable yield losses. Relatively high densities of *A. pisum* were found in habitats dominated by hairy vetch (*Vicia villosa*), which was the only legume other than cultivated dry pea where PEMV was detected. Our results indicate that *V. villosa* is a key alternative host for PEMV, and that pest management practices in this region should consider the distribution and abundance of this weedy host in viral disease mitigation efforts for pulses.

**Keywords:** Aphids, legumes, plant viruses, reservoirs, non-crop hosts, pulse crops

**Introduction**

Plant viruses cause an average 10% reduction in global agricultural productivity, which translates to global economic losses of more than US $30 billion annually (Strange & Scott 2005, Jones 2021). Circulative-transmitted viruses require an insect vector, often phloem-feeding hemipterans like aphids (Power 2000; Hogenhout et al. 2008). Despite the importance of vector-borne plant viruses in agriculture, our ability to predict virus occurrence across time and space remains poor for most pathosystems. Many vectors are generalists with a broad host range that includes food and cover crops, invasive agricultural weeds, and native plants (Mueller et al. 2012, Bommarco et al. 2007). Identifying host reservoirs is key to determine the source(s) of vector-borne pathogens that can outbreak in a crop system (Peterson 2018, Gobatto et al. 2019).

Like their vectors, many crop viruses occupy alternative hosts before infecting crop plants (Norris & Kogan 2005). Non-crop hosts have been established as reservoirs for insect vectors and vector-borne pathogens that infect annual crops such as wheat, corn, and rice (Rashidi et al. 2020, Wu et al. 2020). The replication and spread of a plant virus across multiple hosts depends on the compatibility and coordinated interactions of virus- and host-encoded proteins, and the severity of infection often differs among hosts (Heinlein 2015, Basu et al. 2018). Assessing whether certain hosts act as reservoirs of pathogens can be difficult, especially if alternative hosts do not show clear signs of infection (Lucas 2006, Takahashi et al. 2019). When infection symptoms are not easily observed in the field, management strategies for crop pathogens rely on detection through molecular diagnostic which test plant (Rageshwari et al. 2017). Understanding how pathogens and vectors move among distinct populations of hosts is also an important component of effective management of crop diseases.

When a non-crop host for pathogens or vectors is identified, integrated pest management (IPM) strategies suggest targeted removal to prevent crop infection (Catton et al. 2015, Macharia et al. 2016). For example, management of wheat stem rust relies on control of the pathogen’s alternative host American barberry (*Berberis canadensis*), a strategy that dates back almost a century (Peterson 2018). Non-crop host removal can be difficult if these host plants are also weeds, however, particularly those that emerge early in seasons before crops are established (Norris & Kogan 2005). Consequently, some agricultural weeds and cover crops allow pest insect populations to increase before moving into crops, exacerbating outbreaks (Wenninger et al. 2019). For example, Colorado potato beetle is observed feeding on horse nettle before moving into potato (Mena-Covarrubias et al. 1996), and two spotted spider mites disperse from weeds to cotton (Wilson 1995, Norris & Kogan 2005).

Movement of generalist vectors between non-crop and crop hosts can mediate the spread of pathogens (Power et al. 1991; Davis et al. 2015; Srinivasan et al. 2008). Aphids that migrate over long distances often establish population in crops rapidly in the spring months as non-crop hosts senesce, which is often accompanied by high prevalence of virus-infected plants (Clement et al. 2010, Reynolds et al. 2006). Aphids with long-distance dispersal capability as alates can also complicate management efforts because outbreaks occur at regional scales (Powell et al. 2006, Mueller et al. 2012). For these reasons, aphid-borne viruses are hard to track, and outbreaks are often unpredictable, hampering pest management (Damgaard et al 2020). To address such challenges, identification of local sources of aphid-borne pathogens can be of great value in guiding optimal and cost-effective control strategies, such as removal of weedy reservoirs near crop fields, mitigating damage in outbreak years. To this end, the goal of our study was to track and quantify potential non-crop hosts for a problematic pathogen, PEMV or pea enation mosaic virus, which occurs in our study region of eastern Washington state and Northern Idaho, USA.

**Methods**

***Study System***

The pea aphid *Acyrthosiphon pisum* is a frequent pest of pulse crops that acts as a vector for several economically important pathogens, including PEMV (Rashed et al. 2018; Chatzivassiliou 2021). Plants infected with PEMV produce a range of species-specific symptoms, with malformed pods ultimately reducing yield (Clement et al. 2010). Extreme outbreaks can lead to up to 40% yield loss in pulses (Elbakidze et al. 2011, Paudel et al. 2018). In addition to dry pea (*Pisum sativum*), PEMV infects crops and weeds like alfalfa (*Medicago sativa* L.), yellow sweet clover (*Melilotus officinalis* (L.), white sweet clover (*Melilotus albus* L.), wild white clover (*Trifolium repens* L.), common vetch (*Vicia sativa* L.), hairy vetch (*Vicia villosa* Roth), and broadbean (*Vicia faba* L.) (McEwen et al. 1957). Pea aphids acquire PEMV from a few perennial legume hosts and agricultural weeds (Hull 1981). However, *A. pisum* diet-breadth encompasses most of the Fabaceae (Peccoud et al. 2009), suggesting the diversity of PEMV-compatible hosts could be large.

***Survey Design***

We conducted field surveys from May to July 2018 during an outbreak season of *A. pisum*. Pea aphids and virus have been historically monitored in eastern Washington and Idaho using a long-term trapping network for 17 growing seasons (https://www.legumevirusproject.org). In this trapping scheme, at least ten locations have three pan traps placed at field edges starting after spring peas are planted (May). Pan traps contained propylene glycol for capturing alate aphids, and these were sampled biweekly. Sampling ceased when dry peas in the region complete pod development and are too desiccated to support aphid populations. All pan-trap collected, alate aphids were counted and tested for viral pathogens, including PEMV. In this trapping network, the 2018 season had the second highest alate arrival counts on a per-trap basis over this entire period (Fig. S1). This so-called “outbreak year” thus provided an opportunity to discover the non-crop hosts for *A. pisum* and PEMV in a season when aphids are widespread, thus we targeted sampling at areas with patches of weedy legumes in 60 sites (30 locations >1km apart, each with two repeated visits but samples taken 150m apart). Plant and aphid communities were sampled in two climatic ecoregions: Palouse Prairie, a high-elevation grassland predominately converted to dryland wheat production (Looney and Eigenbrode, 2012) and shrub-steppe, a habitat found at lower elevations and warm slopes adjacent to the Palouse region (predominately along the Snake River in Washington and Idaho) (Knick & Rotenberry 1997). Both habitat types harbor a diverse community of herbaceous legumes and are purported sources of pea aphid outbreaks (Clement 2006). All non-agricultural sites were in either roadside edges, native prairie, or shrub-steppe. Agricultural sites were spring-planted pea fields on the lower Palouse in Whitman Co. Washington and Latah Co. Idaho between 47.46°N and 46.33°N (Fig. 1).

Aphid, plant, and virus surveys were conducted using a line-transect (Fig S2). At each of sites sampled we ran 20m line transects and quantified plant diversity (species identity) of all forbs touching the line transect; forb percent cover was calculated by measuring the length of the line transect (in cm) covered by plant material. At each transect we collected canopy arthropods using two 180° sweeps through the foliage; insects collected were stored in 95% ethanol until identification to species. Samples of aboveground terminal leaf tissue of legume species overlapping the meter-line transect were harvested, wrapped in aluminum foil, frozen in liquid N2, and held on dry ice before storing at -80°C. These tissue samples were used to determine the presence of PEMV.

***PEMV-1 detection in plants by RT-PCR***

To test all crop and non-crop legumes for PEMV, we used a two-stage protocol (Sint et al. 2016). First, we tested for PEMV by using reverse transcription-polymerase chain reaction (RT-PCR) from pooled samples of all tissue collected from each transect (n = 60). Subsamples of tissue from each plant, regardless of species, were pooled and ground into fine powder under liquid N2 by mortar and pestle (Chisholm et al. 2018) and combined into a transect-wide mix. Second, if PEMV was detected in the pooled sample, the remaining tissue from all host plants was tested individually. This method allows efficient scoring of each of plant in a sample for the presence PEMV while avoiding unnecessary and costly sampling of individual plants if the entire population is free of the virus.

For detection of PEMV from plant tissue samples, 100 mg of homogenized tissue was run through Promega SV total RNA isolation kits (Promega), producing cDNA from 1 µg of total RNA using Bio-Rad iScript cDNA synthesis kits (Lee et al. 2021). Then RT-PCR was performed using PEMV-1 coat protein specific primers (PEMV CP FP: 5’ GTGGTGGCACCCTCTATG 3’; PEMV CP RP: 5’ GTGTCCACATGGTAGGCTATG 3’). Primers were designed using the IDT Primer Quest Tool for RT-PCR reaction (10 µl) containing 3 µl of ddH2O, 5 µl of dream Taq mastermix (Thermo Scientific, Waltham, MA, usa), 1 µl of diluted primer mix (forward and reverse [concentration 10µM]), and 1 µl of cDNA template. The RT-PCR program included an initial denaturation for 5 min at 95°C followed by 21 cycles of denaturation at 95°C for 30 s, annealing for 30 s at 56°C, and extension for 45 s at 72°C and final extension of 10 min at 72°C. After PCR was complete, agarose gels (1%) were run at 90 v for 45 min, after which gel pictures were taken in a documentation system (Bio-Rad, Hercules, CA). For one large population of hairy vetch that contained PEMV (Wawawei Park Road, 46.630, -117.378), we revisited the site later in the season and sampled living, adjacent hairy vetch population, validating that PEMV was indeed persistent in this location via collecting. These five additional *V. villosa* samples were processed to rule out contamination as the cause of PEMV detection at this site.

***Statistical Analyses***

All data analyses were completed using R version 4.1.2 (R Development Core Team 2021) using base functions unless otherwise specified. For analyses of plant and aphid data, we used GLMM (generalized linear mixed models) applying the ‘lme4’ package (Bates et al. 2015); model estimates and P-values were extracted using the ‘car’ package (Fox and Weisberg 2011). For plotting results and posthoc tests, we used the ‘emmeans’ package (Lenth 2016). Aphid counts, or cumulative abundance models used a negative-binomial link function appropriate for zero-inflated count data. These abundance data were then transformed for plotting by dividing abundance estimates by total host plant area (Fig. 2). Probability of aphid presence in transects was modeled as the ratio of presence and absence among sites (Fig. 4). Statistical analyses for line transects used ‘site’ as a random effect. Analyses of pooled long-term monitoring data from <https://www.legumevirusproject.org/> were completed using a GLMM with a negative binomial link function. These source data were comprised of samples from a minimum of 30 pan traps monitored weekly over a 17-year survey period (Fig S1).

**Results**

Among all transects, we collected 15,289 *A. pisum* aphids in total and assayed 1,076 candidate plant tissue samples for PEMV. In our transects we recorded 145 species of annual plants, of which 23 were in the family Fabaceae. We observed a range of abundances of aphids on non-crop hosts (Fig. 2) and abundance of non-crop legumes (Fig. 3).

Hairy vetch had the highest abundance of pea aphids and was the most abundant non-crop, weedy legume (Fig. 3). At the community level, increasing coverage of vetch in transects was related to a greater likelihood of pea aphid presence (GLMMs, χ2 = 15.02, *P* < 0.0001, Fig. 4). Notably, adjacent habitats also had high populations of hairy vetch colonized by pea aphids upon subsequent revisit dates (Fig. S3). Finally, PEMV was only detected in hairy vetch (Fig. S4) and crop (dry pea) sites colonized by pea aphids, but not in other hosts.

**Discussion**

Effective prediction of viral plant pathogen outbreaks requires a detailed understanding of vector and pathogen movement from crop to non-crop hosts at the landscape scale (Srinivasan et al. 2008). While our results are limited to a single field season, the first step in risk assessment is evaluating potential alternative hosts during an outbreak (Holt et al. 2008). During an outbreak year for pea aphids, hairy vetch plants were suitable and heavily occupied alternative hosts for pea aphids and is a competent host for PEMV. While other alternative hosts may be found with additional surveys, we found that hairy vetch has high densities in non-agricultural environments, and it is conventionally used in the western U.S. as a cover crop (Luna et al. 2012). Our surveys of plant communities in habitats adjacent to pea fields suggest that there are at least 23 potential hosts that can be resampled in future years, and the absence of aphids or PEMV does not rule them out as compatible hosts for either.

Our understanding of pea aphid and PEMV outbreaks in the Palouse considers that pea aphids likely colonize Palouse agroecosystems following wind currents from the Columbia Basin and Willamette Valley, where milder winters allow aphids to overwinter on alfalfa and clover (Clement et al. 2010, Hampton 1983). Genetic data shows that the pea aphid biotype found on dry pea in the Palouse has shared markers with biotypes collected in these areas (Eigenbrode et al. 2016). Our study suggests two possibilities that align with this information. First, hairy vetch, which emerges early and in low elevation areas that warm up early in the growing season, is an effective “stopover” host for aphid alates dispersing from warmer western regions to the Palouse. Second, hairy vetch occurs in relatively warm microhabitats along the edge of the Palouse and in the lower elevations in the Columbia Basin, and aphids may overwinter in these areas. Vetch is a facultative biennial with an above-ground rosette during ideal climatic conditions (Pokorny et al. 2020, Mischler et al. 2010), and may have a small second generation in mesic habitats in the fall and winter (Clark personal observations). In either case, vetch may act as a short- (months) or long-term (years) reservoir for aphids and PEMV. In some years, PEMV-infected hairy vetch may provide inoculum for arriving aphids, contributing to more injurious infections associated with early transmission to pulse crops (Paudel et al. 2018). Furthermore, in years when infectious aphids arrive later in the season, if they colonize vetch, the pathogen can gain an overwintering foothold for possible infection of legume crops in the following growing season.

Once non-crop hosts for specific plant pathogens are discovered, management implications arise. Removing non-crop host plants could reduce the incidence of that pathogen in crops (Peterson 2018, Strickland et al. 2020). In other systems, management of weeds may reduce pest populations in crops (Norris & Kogan 2005). However, in many cases removal of non-crop hosts may not be viable if they occur over large geographic regions or when movement of pests between hosts occurs over long distances, so local control would not prevent outbreaks. In Palouse agroecosystems it is unclear if a weed removal strategy is tenable. Hairy vetch seeds are spread to produce cattle forage (Golden et al. 2016), possibly explaining how it persists at high abundance in the region. Hairy vetch also improves soil nitrogen, prevents erosion, but it is not listed as a noxious weed in this climatic zone (Pokorny et al. 2020, Mischler et al. 2010). Our results suggest that in this region, cover cropping of hairy vetch may increase PEMV outbreak risk in dry peas if in the same fields, but further work would be needed to verify if region-wide control would reduce likelihood of economic impacts of PEMV.

The optimization of pathogen detection from field samples also depends on precision and specificity of the procedure to ensure efficient and accurate detection of true positive samples (Yazdkhasti et al. 2021). The use of more advanced molecular detection techniques, such as real time-PCR with much lower detection threshold can be used to detect pathogens with low titer (Rubio et al. 2020). Another key step toward improved pathogen detection is to maintain the quality and integrity of field samples by following proper collection technique in order to enhance detection of pathogens from field samples. Based on our observations, Hairy vetch did not exhibit any outward signs of infection by PEMV (e.g. Takahashi et al. 2019), meaning molecular diagnostics would remain necessary to track the spread of these pathogens in this reservoir.

Hairy vetch emerges earlier and hosts pea aphids earlier in the season compared to cultivated legumes. The phenological difference between weeds and crops suggest that a survey of PEMV in vetch may be available to predict seasonal prevalence of PEMV prior to crop emergence. Greenhouse work has demonstrated that pea aphid adults feeding on vetches with PEMV can then transmit these viral pathogens to dry pea (Clark et al. 2019). Sampling hairy vetch for aphids and PEMV may be a way to indicate if there are risks of large-scale, catastrophic outbreaks of PEMV likely to occur each year. While we only have one season of data reported here, PEMV and aphid populations go through large and difficult to predict population cycles, thus more data is necessary to predict population cycles (e.g., Perneck et al. 2008, Northfield et al. 2018). Consequently, it appears likely that if PEMV is found in April or early May in many hairy vetch populations along the lower Palouse, it would portend an areawide impact of PEMV in the growing season for pulse farmers. Similar strategies could be employed in other non-crop and crop source-sink dynamics systems where the non-crop host is a perennial plant that emerges earlier in the growing season.

**Conflict of Interest Statement**

The authors declare no conflicts of interest.

**Author Contribution**

REC, DWC, and SDE and conceived project design. REC and ECO completed surveys and data analysis. REC and SB completed molecular assays. SDE provided long-term aphid trap data. All authors wrote, edited, and approved the final manuscript.

**Data Availability Statement**

All data and R code is available through a publicly available GitHub repository curated by REC at https://github.com/robclark19/vetch2018.

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